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Harmonisation of nucleic acid testing for Zika virus: development of the first WHO International Standard

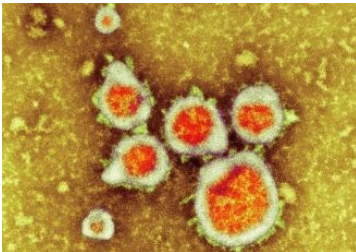
Nucleic acid amplification technique (NAT)-based assays for Zika virus (ZIKV) are important for accurate and timely clinical diagnosis and for screening of blood donors. NAT-based assay results must be reproducible and also consistent between different laboratories performing the same tests. A recent WHO collaborative study to establish an International Standard (IS) for ZIKV RNA concluded that assay variability is reduced when RNA concentrations from both reference strains and clinical samples are reported in relation to the proposed standard: cIS 11468/16. NCPV ZIKV strain 1604131v (strain PRVABC59) was used in the development of this WHO standard.

NCPV is able to supply three ZIKV strains including [NCPV 1604131v](#).

[Find out more about NCPV ZIKV isolates](#)

[Download a copy of the new NCPV brochure!](#)
[Find out about the strains and services available from NCPV.](#)

New strains in the NCPV collection



Respiratory syncytial virus B (RSVB)

[NCPV 1607081v](#), strain RSVB/England653/2015, was sourced from a hospitalised community patient in 2015. The genome has been sequenced although is not currently available on Genbank.

This strain has a genetic shift in the N gene in a specific position which can affect detection assay primers/probe binding. It is therefore useful for molecular assay validation.

[New strains in 2017](#)

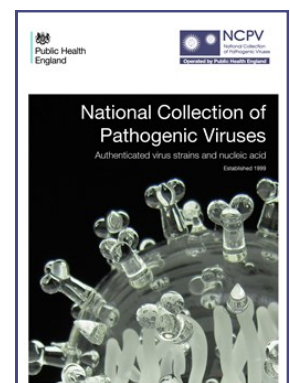
Quality control and authenticity of NCPV strains

Some estimates indicate that up to 70% of microbial strains used in published research are not from recognised culture collections and are used without proper authentication and provenance.

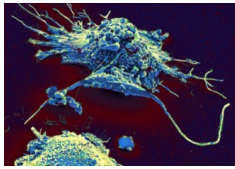
The NCPV team undertakes quality control tests for every batch of virus in the collection including:

- Viability tests – ensures the virus can be grown successfully
- Sterility tests – checks the culture is free from bacterial contamination
- Mycoplasma status – determines whether the cell culture is mycoplasma-free
- PCR – confirms virus identification
- Partial genome sequencing - confirms strain authenticity
- Whole genome sequencing – available for some NCPV viruses

[Download the brochure to find out more](#)



Articles to read.....



The importance of reproducibility in life-science research

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3D cell culture models can be used to study virus-host interactions

[Read the article](#)



Interactions of human microglia cells with Japanese encephalitis virus

[Read the article](#)

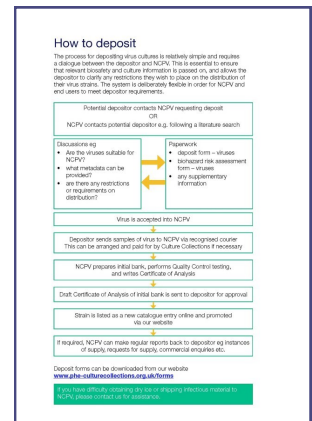
How to deposit with NCPV

The process for depositing virus cultures is relatively simple and requires a dialogue between the depositor and NCPV.

This is essential to ensure that relevant biosafety and culture information is passed onto the NCPV team, and also allows the depositor to clarify any restrictions they wish to place on the distribution of their virus strains.

The system is deliberately flexible so NCPV and end users can meet depositor requirements.

[NCPV deposit flowchart](#)



Meet the team - Lidia Liberty

Lidia is a Healthcare Scientist Supervisor for NCPV. She has worked for PHE for eight years. Her main role involves the production and quality control of viruses (Hazard Group 2 and 3) and overseeing the safe management of day-to-day laboratory operations.

Three facts about Lidia:

- she grew up in Lodz, Poland
- her best holiday was to Greece in 2014, she loved the food, weather and the history
- her guilty pleasure is vanilla ice-cream

Events

NCPV will be attending ECCMID17 come and find us!

27th ECCMID

27th ECCMID 2017

22 - 25 April 2017

Vienna, Austria

- Stand: 34A
- Oral presentation: NCTC_3000 project, 22 April (13:30 - 15:30), Abstract 2956
- Eposter: EV0092



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